Partitioning the influence of soil N, mycorrhizae, and foliar N uptake on foliar δ¹⁵N patterns

D.M. Vallano and J.P. Sparks

e-mail: dvallano@ucsc.edu

Department of Ecology and Evolutionary Biology. Cornell University



Introduction

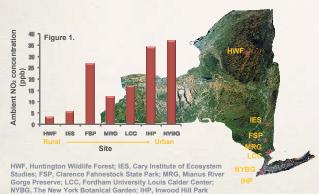
- Vegetation is an important sink for atmospheric reactive N in N limited systems and direct foliar uptake may be a significant pathway for the acquisition of plant-available N.
- A proxy for atmospheric reactive N would be useful to estimate the magnitude of this uptake pathway.
- N stable isotope ratios (¹⁵N/¹⁴N) are useful tools because plant-available N sources often have different isotopic N compositions (δ¹⁵N). However, the mechanisms driving differences in foliar δ¹⁵N patterns are still unresolved.
- Foliar δ¹⁵N primarily reflects the integration of soil solution δ¹⁵N, direct foliar N uptake, within-plant fractionations, and fractionation due to mycorrhizae.

Objective

Estimate the influence of soil $\delta^{15}N$, mycorrhizae, and foliar N uptake on foliar $\delta^{15}N$ patterns of several tree species along a gradient of increasing atmospheric NO₂ concentration in New York State (Fig. 1)

We addressed these questions using two approaches:

- 1. Estimate the influence of mycorrhizal fractionation on foliar $\delta^{15}N$ in red maple seedlings using a *potted plant study* where the influence of mycorrhizae was eliminated using fungicide
- Field measurements of foliar and soil δ¹⁵N along a gradient of atmospheric nitrogen dioxide (NO₂) concentration



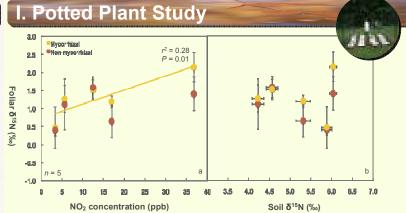


Figure 2. Relationship between foliar $\delta^{15}N$ and average atmospheric NO_2 concentration (a) and bulk soil $\delta^{15}N$ (b) in red maple seedlings with (yellow) and without (red) mycorrhizae. We observed a positive correlation between foliar $\delta^{15}N$ and increasing NO_2 concentration in red maple seedlings with mycorrhizae, but when mycorrhizae were inhibited with fungicide this relationship no longer existed. Also, the presence of mycorrhizae did not significantly alter foliar $\delta^{15}N$. Data are means \pm 1 SE.

Figure 3. Relationship between foliar and bulk soil $\delta^{15}N$ across a gradient of atmospheric NO $_2$ concentration in adult trees (left panels) and the partial residual plots for the predictor variables included in the multiple regression model predicting foliar $\delta^{15}N$ (right panels). We found strong relationships among soil $\delta^{15}N$, NO $_2$ concentration, and foliar $\delta^{15}N$ with varying strength across species (red maple > red oak > American beech > black birch). Multiple regression analysis revealed that for some species the average NO $_2$ concentration explained significant additional variation after site level bulk soil $\delta^{15}N$ was accounted for in the model. Data are means \pm 1 SE (n = 20).

III. Evidence for foliar N uptake?

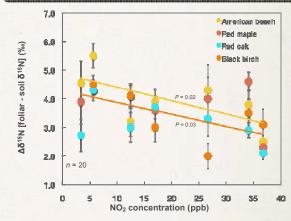


Figure 4. Relationship between $\Delta \delta^{15}N$ (absolute difference between soil and foliar $\delta^{15}N$) and atmospheric NO₂ concentration. Data are means \pm 1 SE. The *a priori* prediction was a divergence between soil and foliar $\delta^{15}N$ would increase at sites with a higher potential for foliar uptake. However, this was not the observation. Soil $\delta^{15}N$ tended to increase across the gradient (i.e., driving this relationship in the opposite direction). There are two other possibilities: (1) even though soil $\delta^{15}N$ cannot explain the trend, differential rates of mineralization/nitrification across the gradient may lead to differential soil solution $\delta^{15}N$ helping to drive this pattern, and (2) fractionation events influencing ^{15}N during NO₂ assimilation (diffusion, enzymatic fixation, etc.) are unknown. Thus, the influence of foliar uptake in this context is poorly constrained and, depending upon the $\delta^{15}N$ of the pathway, could generate this pattern.

Conclusions

- Effect of mycorrhizae on foliar δ¹⁵N minimal in this system
- Results suggest direct foliar N uptake of atmospheric reactive N deposition has an influence on foliar δ¹⁵N in several species and can be detected using natural abundance δ¹⁵N measurements of plant and soil material.

Acknowledgement

This research was funded by a Kieckhefer Adirondack Fellowship and by the Cornell NSF-IGERT in Biogeochemistry and Environmental Biocomplexity, Award #DGE 0221658. We thank Jocelyn Lavallae, Rebecca Doyle-Morin and Danica Lombardozzi for field support. Special thanks to those who have assisted us with this project at the following sites: Adirondack Ecological Center, Fordham University's Louis Calder Center, the Cary Institute for Ecosystem Studies, Mianus River Gorge Preserve, Clarence Fahnestock State Park, The New York Botanical Garden, and Invood Hill Park.